

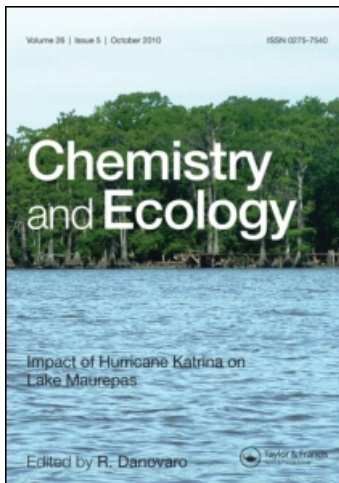
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## Chemistry and Ecology

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**To cite this Article** Hwang, Deng-Fwu, Lin, Jui-Fen and Jeng, Sen-Shyong(1996) 'Comparative Toxicity of Copper and Zinc to Isolated Eel Hepatocytes', *Chemistry and Ecology*, 12: 1, 109 – 114

**To link to this Article:** DOI: 10.1080/02757549608035351

**URL:** <http://dx.doi.org/10.1080/02757549608035351>

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## COMPARATIVE TOXICITY OF COPPER AND ZINC TO ISOLATED EEL HEPATOCYTES

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(Received 24 August 1995; Revised 29 November 1995)

Attempts were made to elucidate lethal responses of hepatocytes of Japanese eel (*Anguilla japonica*) exposed to Krebs-HEPES buffer at different concentrations of  $\text{Cu}^{++}$  and  $\text{Zn}^{++}$ . It was found that the 2-hr  $\text{LC}_{50}$  value of  $\text{Cu}^{++}$  and  $\text{Zn}^{++}$  on eel hepatocytes was 490 and 1576  $\mu\text{M}$  for  $\text{Cu}^{++}$  and  $\text{Zn}^{++}$  in single solution, and 1734 and 5200  $\mu\text{M}$  in mixed solution. The toxic effects of  $\text{Cu}^{++}$  and  $\text{Zn}^{++}$  on eel hepatocytes were characteristic of antagonistic interaction. After 2-hr exposure, the amount of accumulated metal on the hepatocytes and the mortality of hepatocytes were increased with increasing metal concentration in Krebs-HEPES buffer. The accumulated amount of both metals was significantly decreased when the eel hepatocytes were exposed to mixed metals than to single metal.

KEY WORDS: Copper, zinc, Japanese eel hepatocytes, toxicity,  $\text{LC}_{50}$ .

### INTRODUCTION

Recently, the use of isolated fish hepatocytes as a model system for general toxicity studies has been developed and reviewed (Baksi and Frazier, 1990). Denizeau and Marion (1990) investigated the effects of cadmium, mercury and copper, either in single or in various binary combination, on isolated rainbow trout (*Onchorynchus mykiss*) hepatocytes. They found that cadmium and mercury in combination produced a synergistic interaction while other combinations did not demonstrate interaction. The induction of metallothionein-like proteins by cadmium in isolated striped bass (*Morone saxatilis*) hepatocytes was investigated (Baksi and Frazier, 1988). Ballatori *et al.* (1988) reported the effects of various mercurials and cadmium chloride on plasma membrane permeability of isolated skate (*Raja erinacea*) hepatocytes, and found that plasma membrane was a sensitive target for metal induced toxicity. Rainbow trout hepatocytes were sensitive to aflatoxin  $\text{B}_1$  and acetaminophen as assessed by lactate dehydrogenase leakage (Kocal *et al.*, 1988).

In West Taiwan, copper and zinc are often reported as pollutants in aquaculture area. Although there have many reports on the toxicity of copper and zinc on fish (Taylor *et al.*, 1985; Elsa, 1991), toxic effects of both pollutants on fish hepatocytes are not reported so far. Because Japanese eel (*Anguilla japonica*) is an important species for aquaculture in western Taiwan, the study of acute toxic effects of copper and zinc on the eel hepatocytes was undertaken to evaluate concentration-response relationship and to know the mode of interaction between copper and zinc.

## MATERIALS AND METHODS

### *Animals*

Japanese cultured eels, *Anguilla japonica*, weighing from 350–450 g, were purchased from a local aquaculturist in Ilan Prefecture. They were kept in freshwater aquaria and fasted for about three days before use.

### *Chemicals*

Tissue culture reagents were purchased from Sigma Chemical Co. (St. Louis, MD, USA). Copper chloride, zinc sulphate and other reagents were from E. Merck & Co. (Darmstadt, Germany).

### *Hepatocyte Isolation*

The eels were anaesthetized with 0.3% ethylene glycol monophenol ether. The liver was isolated and perfused as described previously (Hwang *et al.*, 1994). Briefly, the eel livers were continuously perfused with Hanks buffer, Hanks EGTA buffer, Hanks enzyme buffer, and Krebs-HEPES buffer, and then suspended with Krebs-HEPES buffer. The viability of suspended hepatocytes was determined by trypan blue dye exclusion.

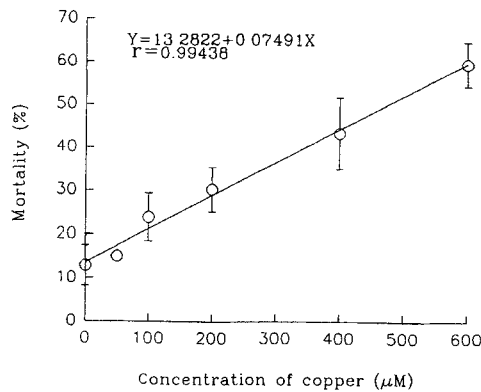
### *Test for Toxicity of Copper and Zinc*

The suspended hepatocytes were diluted with Krebs-HEPES buffer to a cell concentration of  $2 \times 10^7$  cells/ml into tubes. The diluted hepatocytes solution in each tube was separately exposed to different copper (as  $\text{CuCl}_2$  form) concentrations [control ( $\text{Cu}^{++}$ -free), 50, 100, 200, 400 and 600  $\mu\text{M}$   $\text{Cu}^{++}$ ], zinc (as  $\text{ZnSO}_4$  form) concentrations [control ( $\text{Zn}^{++}$ -free), 50, 100, 500, 1000 and 2000  $\mu\text{M}$   $\text{Zn}^{++}$ ], and  $\text{Cu}^{++}$ - $\text{Zn}^{++}$  mixed concentrations [control ( $\text{Cu}^{++}$ - $\text{Zn}^{++}$ -free, 200  $\mu\text{M}$   $\text{Cu}^{++}$ -600  $\mu\text{M}$   $\text{Zn}^{++}$ , 400  $\mu\text{M}$   $\text{Cu}^{++}$ -1200  $\mu\text{M}$   $\text{Zn}^{++}$ , 800  $\mu\text{M}$   $\text{Cu}^{++}$ -2400  $\mu\text{M}$   $\text{Zn}^{++}$ , 1200  $\mu\text{M}$   $\text{Cu}^{++}$ -3600  $\mu\text{M}$   $\text{Zn}^{++}$ , 1600  $\mu\text{M}$   $\text{Cu}^{++}$ -4800  $\mu\text{M}$   $\text{Zn}^{++}$ , and 2000  $\mu\text{M}$   $\text{Cu}^{++}$ -6000  $\mu\text{M}$   $\text{Zn}^{++}$ ]. After 2-hr exposure, the solution in the tube was pipetted vigorously to mix and then number of the hepatocytes of the suspension obtained was counted by a blood counter. The experiment for each test was determined in triplicate. The data on hepatocyte viability were collected following methods described by Buikema *et al.* (1982), Trevors and Lusty (1985), and Roy (1988) to provide 2-hr median lethal concentration ( $\text{LC}_{50}$ ) values and the associated 95% confidence intervals (CI). The above pairing ratio of  $\text{Zn}^{++}$  to  $\text{Cu}^{++}$  was three; this ratio was obtained from 2-hr  $\text{LC}_{50}$  value of  $\text{Zn}^{++}$  to that of  $\text{Cu}^{++}$ . On the other hand, 1 ml of hepatocyte solution from each test tube was removed and centrifuged at 500 rpm for 5 min. The precipitated hepatocytes were determined for metal content by using a flame atomic absorption spectrophotometer (Hitachi Z-8100, Hitachi Co., Japan) (Apha, 1985).

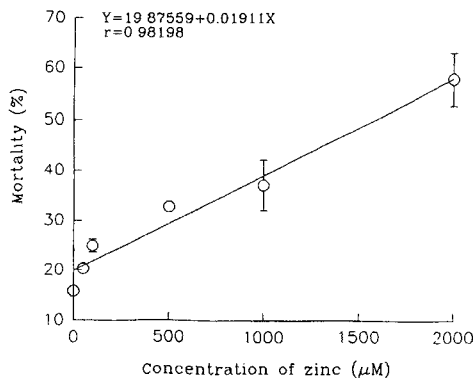
## RESULTS

When the eel hepatocytes were exposed to either  $Zn^{++}$  or  $Cu^{++}$  in single solution for 2 hr, the relationships between mortality and concentration of both metals are shown in Figures 1 and 2. There are two positive linear relationships for  $Y = 13.28 + 0.075 X$  ( $r = 0.99$ ) and  $Y = 19.87 + 0.019 X$  ( $r = 0.98$ ) for  $Cu^{++}$  and  $Zn^{++}$ , respectively. The eel hepatocytes demonstrated more sensitivity to  $Cu^{++}$  than to  $Zn^{++}$ . The 2-hr  $LC_{50}$  values and their 95% confidence limits for  $Cu^{++}$  and  $Zn^{++}$  on eel hepatocytes were  $490 \mu M$  and  $227-753 \mu M$  for  $Cu^{++}$ , and  $1576 \mu M$  and  $904-2248 \mu M$  for  $Zn^{++}$ . This indicated that 2-hr  $LC_{50}$  value of  $Zn^{++}$  was about three fold that of  $Cu^{++}$  for eel hepatocytes.

The relationships between mortality and mixed concentration of  $Zn^{++}$  and  $Cu^{++}$  for eel hepatocytes exposed for 2 hr are shown in Figure 3. There are also two positive linear relationships for  $Y = 15.15 + 0.0067 X$  and  $Y = 15.15 + 0.02 X$  ( $r = 0.98$ ) for  $Zn^{++}$  and  $Cu^{++}$ , respectively. The respective  $LC_{50}$  value of  $Zn^{++}$  and  $Cu^{++}$  was  $5200 \mu M$  and  $1734 \mu M$ . Their 95% confidence limits for  $Zn^{++}$  and  $Cu^{++}$  on eel hepatocytes



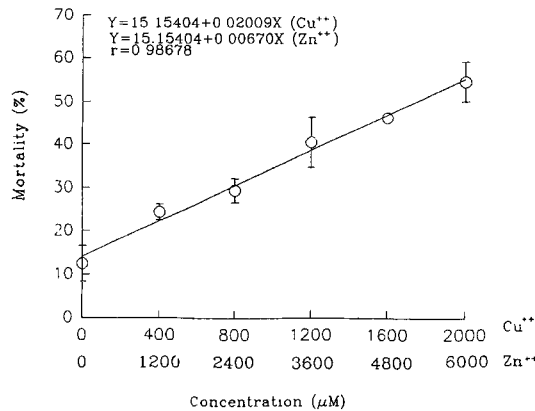
**Figure 1** Relationships between mortality and concentration of copper on eel hepatocytes exposed for 2 hr.



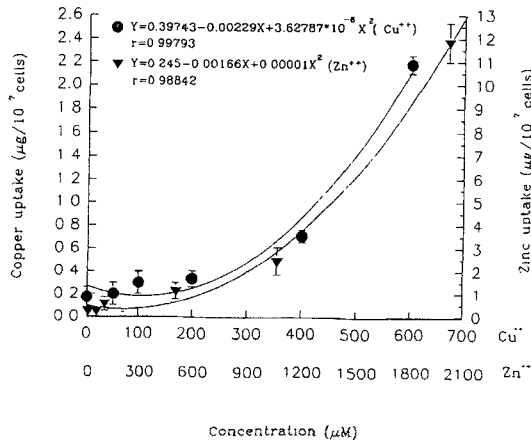
**Figure 2** Relationships between mortality and concentration of zinc on eel hepatocytes exposed for 2 hr.

were 3617–6784 and 1206–2263  $\mu\text{M}$ , respectively. The 2-hr  $\text{LC}_{50}$  values of  $\text{Zn}^{++}$  and  $\text{Cu}^{++}$ , when eel hepatocytes were exposed to  $\text{Zn}^{++}$ - $\text{Cu}^{++}$  mixed solution, were about 3.5 fold those when the hepatocytes were separately exposed to  $\text{Zn}^{++}$  or  $\text{Cu}^{++}$  solution. This indicate the toxic effects of  $\text{Zn}^{++}$  and  $\text{Cu}^{++}$  on eel hepatocytes are antagonistic interaction.

When the eel hepatocytes were exposed to  $\text{Zn}^{++}$  and  $\text{Cu}^{++}$  in single solution, accumulated amounts of  $\text{Zn}^{++}$  and  $\text{Cu}^{++}$  on the eel hepatocytes are shown in Figure 4. It was found that amounts of accumulated of both metals were increased with increasing the concentration of exposed metal, especially when the concentration of  $\text{Cu}^{++}$  and  $\text{Zn}^{++}$  in the buffer was more than 200  $\mu\text{M}$  and 500  $\mu\text{M}$ , respectively. The relationships between accumulated and exposed concentrations of metals were as follows:  $Y = 0.40 - 0.0023 X + 3.63 \times 10^{-6} X^2$  ( $r = 0.99$ ) for  $\text{Cu}^{++}$  and



**Figure 3** Relationships between mortality and concentration of copper and zinc on eel hepatocytes exposed for 2 hr.



**Figure 4** Copper and zinc uptake in eel hepatocytes incubated in Krebs-HEPES buffer (pH 7.5) with different concentration of single metal for 2 hr.

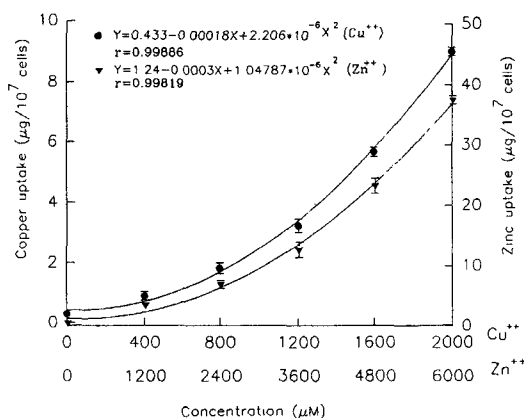
$Y = 0.24 - 0.0017 X + 0.00001 X^2$  ( $r = 0.99$ ) for  $Zn^{++}$ , respectively. The pattern of relationships between accumulated and exposed concentrations of both metals in the eel hepatocytes were rather similar to each other. It means that both metals might have the same passway to enter into eel hepatocytes.

When the eel hepatocytes were exposed to  $Cu^{++} - Zn^{++}$  mixed solutions, the accumulated amounts of  $Cu^{++}$  and  $Zn^{++}$  on the eel hepatocytes are shown in Figure 5. It was also found that amounts of both accumulated metals were increased with increasing the concentration of exposed metal. The amounts accumulated of both metals in eel hepatocytes were less in mixed solution than in single solution. The relationships between accumulated and exposed concentrations of metals were as follows:  $Y = 0.43 - 0.00018 X + 2.20 \times 10^{-6} X^2$  ( $r = 0.99$ ) for  $Cu^{++}$  and  $Y = 1.24 - 0.0003 X + 1.047 \times 10^{-6} X^2$  ( $r = 0.99$ ) for  $Zn^{++}$  respectively.

## DISCUSSION

The responses of Japanese eel hepatocytes exposed to lethal level of  $Zn^{++}$  and  $Cu^{++}$  solutions were demonstrated in this study. It was found that the respective 2-hr  $LC_{50}$  value of  $Cu^{++}$  and  $Zn^{++}$  on eel hepatocytes was 490 and 1576  $\mu M$  for  $Cu^{++}$  and  $Zn^{++}$  in single solution, and 1734 and 5200  $\mu M$  in mixed solution. The toxic effect of  $Cu^{++}$  on Japanese eel hepatocytes was higher than that of  $Zn^{++}$ . This phenomenon was the same as that on other fish (Thompson *et al.*, 1980, 1980; Taylor *et al.*, 1985; Cusimano and Brakke, 1986). Moreover, the toxic effects of  $Cu^{++}$  and  $Zn^{++}$  on Japanese eel hepatocytes were characteristic of antagonistic interaction. However, the toxic effects of  $Cu^{++}$  and  $Zn^{++}$  on fish were synergistic and/or additive interactions (Lloyd, 1961; Sprague and Ramsay, 1965; Lewis, 1978; Thompson *et al.*, 1980; Elsa, 1991; Alabaster *et al.*, 1994).

No matter how Japanese eel hepatocytes were exposed in  $Cu^{++}$  and/or  $Zn^{++}$  solutions, the amount of accumulated metal on the hepatocytes and the mortality of



**Figure 5** Copper and zinc uptake in eel hepatocytes incubated in Krebs-HEPES buffer (pH 7.5) with different concentrations of mixed metals for 2 hr.

hepatocytes were increased with increasing the concentration of exposed metal. It means that the death of eel hepatocytes might be caused by increasing accumulated amounts of  $\text{Cu}^{++}$  and  $\text{Zn}^{++}$  on the hepatocytes. The amounts of both accumulated metals were significantly decreased when the Japanese eel hepatocytes were exposed to mixed solution than to single solution. This phenomenon might elucidate why the toxic effects of  $\text{Cu}^{++}$  and  $\text{Zn}^{++}$  on the Japanese eel hepatocytes were antagonistic interaction. The mechanism of antagonistic interaction for the toxic effects of  $\text{Cu}^{++}$  and  $\text{Zn}^{++}$  on the eel hepatocytes is now being studied.

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